TECHNICAL NOTE



## The complete mitochondrial genome of *Dosinia japonica* (Bivalvia: Veneridae)

Changda Lv<sup>1</sup> · Lingfeng Kong<sup>1</sup> · Hong Yu<sup>1</sup> · Qi Li<sup>1</sup>

Received: 10 July 2017 / Accepted: 8 August 2017 / Published online: 18 August 2017 © Springer Science+Business Media B.V. 2017

**Abstract** *Dosinia japonica* is a marine bivalve mollusk of the family Veneridae, which is distributed in coastal areas of China, Korean peninsula, Japan, and Russia. In this study, the complete mitochondrial genome of the clam *D. japonica* was determined firstly. It is 17,693 bp in length, including 13 protein-coding genes, 2 ribosomal RNAs, 22 transfer RNAs, and a major non-coding control region. Phylogenetic analyses indicated that *D. japonica* was the sister group of Tapetinae clade, which means Dosininae and Tapetinae have a common ancestor. This report provides molecular biology data for the conservation and management of *D. japonica* in future.

**Keywords** *Dosinia japonica* · Veneridae · Mitochondrial genome · Conservation

The bivalve *Dosinia japonica* belongs to the family Veneridae, which inhabits from intertidal zone to muddy sediments of the shallow coastal (Liu et al. 2010). It distributes in coastal areas of China, Korean peninsula, Japan, and the Russian Far East. In recent years, the natural population of *D. japonica* have declined remarkably due to over-exploitation (Zhang et al. 2013). Therefore, it is necessary to create a valid conservation strategy. Now, there is very little molecular information about this species, while the complete mitochondrial DNA sequences could be informative at deep phylogenetic levels (Curole and Kocher 1999). In this study, we sequenced and annotated the complete mitochondrial

☑ Qi Li qili66@ouc.edu.cn genome of *D. japonica* firstly, and examine the phylogenetic position of *D. japonica* within Veneridae to provide the basic data for conservation genetics.

A live individual of *D. japonica* sample was collected from the coastal water of Ganyu, Jiangsu Province, China. The total genomic DNA was extracted from adductor muscle by a modification of the standard phenol–chloroform procedure described by (Li et al. 2002). The complete mitochondrial genome of *D. japonica* was sequenced using long-PCR technique. All sequence data were sequenced and annotated following the methods described by Yuan et al. (2012).

The complete mt genome of *D. japonica* is 17,693 bp in length (GenBank accession no. MF401432) containing 13 protein-coding genes (PCGs), 2 rRNA genes and 22 tRNA genes (Fig. 1). All 37 genes of *D. japonica* were encoded on the (+) strand. The overall base composition for the mt genome sequence of *D. japonica* was as follows: A=28.21%, C=9.12%, G=20.91%, T=41.76%. The A+T content of the PCGs, rRNAs, tRNAs and non-coding regions (NCRs) was 68.2%, 72.02%, 72.23% and 74.56%, giving a total A+T content of 69.97% in the whole mitochondrial genome. The nucleotide compositions of the entire *D. japonica* mtDNA indicate the occurrence of more T and A than G and C (AT skew, -0.1936; GC skew, 0.3926).

Total length of the 13 protein coding genes is 11,478 bp, eight of the 13 protein coding genes start with ATA or ATG, meanwhile, *co2*, *nad2*, *nad3* genes start with the altercative start codon GTG, and the *Cytb* gene starts with a ATT codon. Eight genes are terminated by TAA, but *cox1*, *nad1*, *cox2*, *nad4* and *cox3* by TAG (Table 1).

The *rrnL* and *rrnS* genes are 1203 and 902 bp in length, respectively. There are 22 typical tRNA genes in the mt genome of *D. japonica*, which range from 61 to 68 bp. The mt genome of *D. japonica* contained 25 non-coding regions ranging in size from 1 to 2027 bp. The total length of all

Key Laboratory of Mariculture, Ministry of Education, Ocean University of China, Qingdao 266003, China

Phylogenetic trees were reconstructed using Maximumlikelihood (ML) analysis and Bayesian inference (BI) approaches. The ML analysis was conducted with PhyML 3.0 (Guindon et al. 2010) and 100 bootstraps were used to assess the support of nodes. BI was performed using MrBayes 3.1.2 (Ronquist and Huelsenbeck 2003). The topological structure showed that the results of ML and BI trees were in almost complete agreement (Fig. 2). This indicates that D. japonica formed a single branch as the sister group of Tapetinae clade, which is consistent with the findings of Chen et al. (2011).

Fig. 1 Complete mitochondrial genome of Dosinia japonica

non-coding regions was 2708 bp, accounting for 15.31%

of the complete mtDNA. The major non-coding region is

2027 bp, which located between *tRNA<sup>lle</sup>* and *tRNA<sup>Asp</sup>* with an

AT content of 54%. The major non-coding region included a

75 bp repeat motif, and one point nine copies were found in

the family Veneridae, the nucleotide sequences of 12

concatenated protein-coding genes (except atp8 gene) of

To identify the taxonomic position of D. japonica in

it (location: 12,654–12,728, and 12,729–12,792).



japonica

| Gene                | From   | То     | Size (bp) | Start | Stop | Intergenic<br>nucleotide |
|---------------------|--------|--------|-----------|-------|------|--------------------------|
| coxl                | 1      | 1608   | 1608      | GTG   | TAG  | 37                       |
| $tRNA^{Leu(CUN)}$   | 1619   | 1684   | 66        |       |      | 10                       |
| nad1                | 1702   | 2607   | 906       | ATG   | TAG  | 17                       |
| nad2                | 2695   | 3714   | 1020      | GTG   | TAA  | 87                       |
| nad4l               | 3717   | 3992   | 276       | ATA   | TAA  | 2                        |
| cox2                | 4071   | 5081   | 1011      | GTG   | TAG  | 78                       |
| tRNA <sup>Pro</sup> | 5086   | 5150   | 65        |       |      | 4                        |
| cytb                | 5173   | 6363   | 1191      | ATT   | TAA  | 22                       |
| rrnL                | 6364   | 7566   | 1203      |       |      | 0                        |
| atp8                | 7567   | 7680   | 114       | ATG   | TAA  | 0                        |
| nad4                | 7684   | 9042   | 1359      | ATG   | TAG  | 3                        |
| tRNA <sup>His</sup> | 9042   | 9103   | 62        |       |      | -1                       |
| tRNA <sup>Glu</sup> | 9106   | 9166   | 61        |       |      | 2                        |
| $tRNA^{Ser(UCN)}$   | 9164   | 9226   | 63        |       |      | -3                       |
| atp6                | 9227   | 9955   | 729       | ATG   | TAA  | 0                        |
| nad3                | 9980   | 10,384 | 405       | GTG   | TAA  | 24                       |
| nad5                | 10,385 | 11,887 | 1503      | ATA   | TAA  | 0                        |
| tRNA <sup>lle</sup> | 12,122 | 12,186 | 65        |       |      | 234                      |
| $tRNA^{Asp}$        | 14,214 | 14,279 | 66        |       |      | 2027                     |
| $tRNA^{Tyr}$        | 14,327 | 14,388 | 62        |       |      | 47                       |
| nad6                | 14,429 | 14,914 | 486       | ATG   | TAA  | 40                       |
| $tRNA^{Lys}$        | 14,919 | 14,984 | 66        |       |      | 4                        |
| $tRNA^{Val}$        | 14,986 | 15,047 | 62        |       |      | 1                        |
| tRNA <sup>Phe</sup> | 15,053 | 15,114 | 62        |       |      | 5                        |
| $tRNA^{Trp}$        | 15,121 | 15,183 | 63        |       |      | 6                        |
| tRNA <sup>Arg</sup> | 15,184 | 15,244 | 61        |       |      | 0                        |
| $tRNA^{Leu(UUR)}$   | 15,245 | 15,309 | 65        |       |      | 0                        |
| $tRNA^{Gly}$        | 15,329 | 15,390 | 62        |       |      | 15                       |
| tRNA <sup>Gln</sup> | 15,391 | 15,457 | 67        |       |      | 0                        |
| tRNA <sup>Asn</sup> | 15,468 | 15,534 | 67        |       |      | 10                       |
| $tRNA^{Thr}$        | 15,543 | 15,604 | 62        |       |      | 8                        |
| rrnS                | 15,605 | 16,506 | 902       |       |      | 0                        |
| tRNA <sup>Met</sup> | 16,507 | 16,574 | 68        |       |      | 0                        |
| $tRNA^{Cys}$        | 16,592 | 16,655 | 64        |       |      | 17                       |
| $tRNA^{Ser(AGN)}$   | 16,660 | 16,726 | 67        |       |      | 4                        |
| cox3                | 16,727 | 17,596 | 870       | ATG   | TAG  | 0                        |
| tRNA <sup>Ala</sup> | 17,595 | 17,656 | 62        |       |      | -2                       |

Fig. 2 Phylogenetic trees of *Dosinia japonica* based on the nucleotide sequences of 12 concatenated protein-coding genes, using *Solen grandis* and *Solen strictus* as the outgroup taxa. The first number at each node is Bayesian posterior probability and the second number is ML bootstrap values



The complete mitochondrial genome sequenced here will be useful for population genomic studies of *D. japonica*, which will play an important role in formulating new conservation and management strategies.

Acknowledgements This study was supported by the grants from Fundamental Research Funds for the Central Universities (201762014), Shandong Province (2016ZDJS06A06), and Shandong Seed Project.

## References

- Chen J, Li Q, Kong LF, Zheng XD (2011) Molecular phylogeny of venus clams (Mollusca, Bivalvia, Veneridae) with emphasis on the systematic position of taxa along the coast of mainland China. Zool Scr 40:260–271. doi:10.1111/j.1463-6409.2011.00471.x
- Curole JP, Kocher TD (1999) Mitogenomics: digging deeper with complete mitochondrial genomes. Trends Ecol Evol 14:394–398. doi:10.1016/s0169-5347(99)01660-2

- Guindon S, Dufayard JF, Lefort V, Anisimova M, Hordijk W, Gascuel O (2010) New algorithms and methods to estimate maximumlikelihood phylogenies: assessing the performance of PhyML 3.0. Syst Biol 59:307–321. doi:10.1093/sysbio/syq010
- Li Q, Park C, Kijima A (2002) Isolation and characterization of microsatellite loci in the Pacific abalone, *Haliotis discus hannai*. J Shellfish Res 21:811–815
- Liu Z, Li J, Yang X, Wang H, Li Y, Zhang S, Xu F (2010) Species of common *Dosinia* from intertidal zones of northern China. Marine Sci 34:30–35
- Ronquist F, Huelsenbeck JP (2003) MrBayes 3: Bayesian phylogenetic inference under mixed models. Bioinformatics 19:1572–1574. doi:10.1093/bioinformatics/btg180
- Yuan Y, Li Q, Yu H, Kong L (2012) The complete mitochondrial genomes of six heterodont bivalves (Tellinoidea and Solenoidea): variable gene arrangements and phylogenetic implications. PLoS ONE 7:e32353
- Zhang W, Chang Y, Ding J, Xiao D, Yu J, Zhou H (2013) Correlation and regression of shell size and weight of *Dosinia japonica* Reeve. Oceanol Limnol Sin 44:796–800